

Design And *In Silico* Validation Of PCR-metabarcoding Primers

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First of all: system settings

```
> ssh username@milou.uppmax.uu.se
> interactive -n X -t X:00:00 -A g2016021
> cp -r /proj/g2016021/metabarcoding/
> cd glob/metabarcoding
> module load python/2.7.9
> python get-obitools.py
> ./obitools
> exit
> tar -zxvf ecoPCR.tar.gz
> cd ecoPCR/src/
> make
> export PATH=$PATH:/proj/g2016021/metabarcoding/ecoPCR/src
> cd ../../..
> tar -zxvf ecoPrimers.tar.gz
> cd ecoPrimers/src/
> make
> export PATH=$PATH:/proj/g2016021/metabarcoding/ecoPrimers/src
```

PROPERTIES

→ Amplify a suitable marker:

- 1) Mutation rate: distinguish species.
- 2) Conserved regions: universal primers.
- 3) Appropriate length: variation without loss of information when degraded.
- 4) Reference libraries: taxonomic identification.

→ Amplify sequences of ALL the species belonging to the target taxon present in the sample.

- 1) Ideally, amplify sequences of NONE of the species NOT belonging to the target taxon present in the sample. This can be a secondary (eDNA) or principal (dietDNA) requisite.
- 2) Amplify all sequences EQUITATIVELY = no amplification bias.

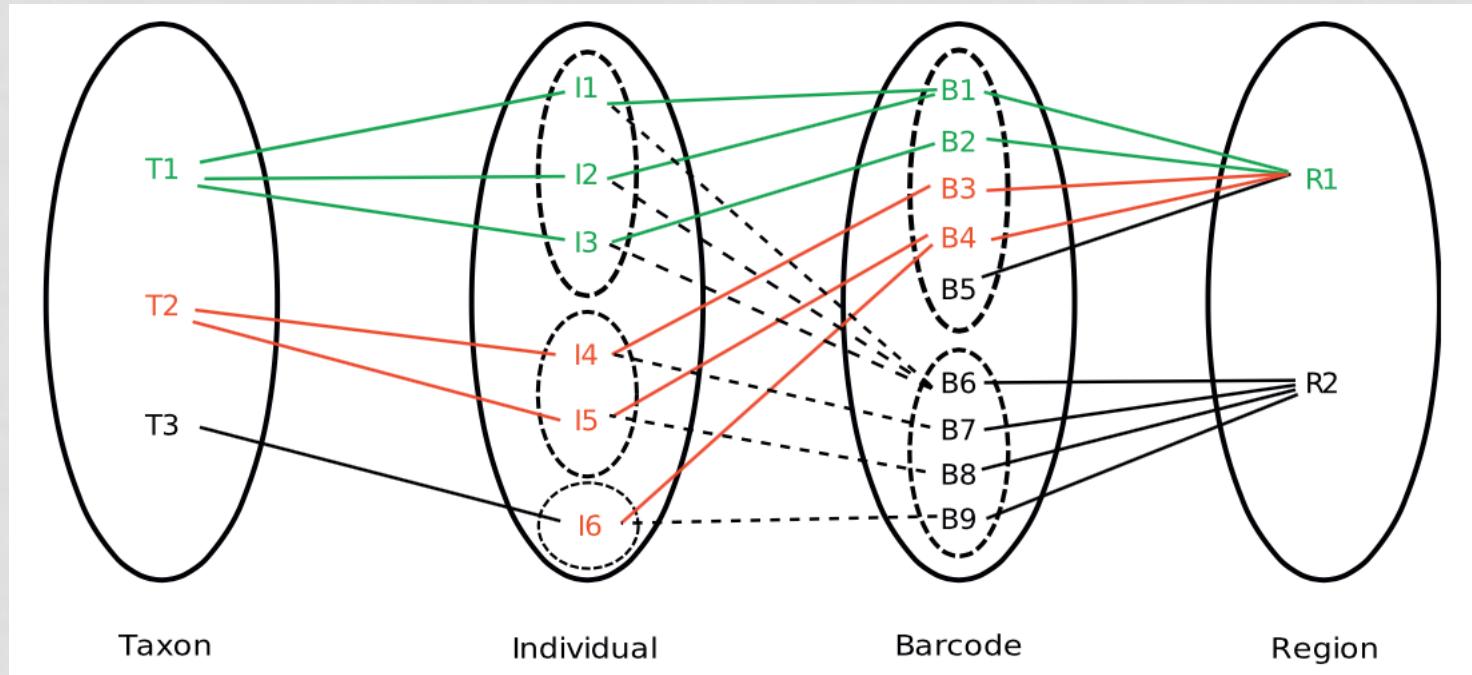
→ Region amplified of the ‘suitable marker’ should discriminate between closely related species.

Properties - Indexes

Bc: Taxonomic coverage $Bc = \text{no. sequences amplified} / \text{no. sequences present}$
PRIMER PROPERTY $Bc = [0,1]$

Bs: Resolution capacity $Bs = \text{no. taxa unambiguously identified} / \text{no. sequences amplified}$

BARCODE PROPERTY $Bs = [0,1]$



ecoPrimers

(Riaz *et al.* 2011)

- Different software from OBITools, but in the same package.
- Specifically developed for metabarcoding (of any taxonomic group).
- Python based.
- Algorithm: Strict Primer Algorithm (SPA).
- Gives nice output.

```
# ecoPrimer version 0.3
# Rank level optimisation : species
# max error count by oligonucleotide : 3
#
# Restricted to taxon:
#   6960 : Hexapoda (superclass)
#
# strict primer quorum : 0.70
# example quorum : 0.90
# counterexample quorum : 0.10
#
# database : sixlegs
# Database is constituted of 1602 examples corresponding to 1115 species
#           and 0 counterexamples corresponding to 0 species
#
# amplifiat length between [50,500] bp
# DB sequences are considered as circular
# Pairs having specificity less than 0.60 will be ignored
#
  0 ATAGAAACCAACCTGGCT    TTACCTTAGGGATAACAG  53.6  1.7  47.7  27.0  8   7   GG   1514  0   0.945  1059  0   0.950  835   0.788  138   217   142.67  #165
  1 ATAGAAACCAACCTGGCT    TACCTTAGGGATAACAGC  53.6  1.7  50.6  30.9  8   8   GG   1502  0   0.938  1048  0   0.940  824   0.786  137   216   141.67  #165
  2 GATAGAAACCAACCTGGC    TACCTTAGGGATAACAGC  53.6  2.0  50.6  30.9  9   8   GG   1499  0   0.936  1046  0   0.938  822   0.786  138   217   142.67  #165
  3 ATAGAAACCAACCTGGCT    GACTCTGATTTGGATTA  53.6  11.4 51.8   38.1  8   8   GG   1498  0   0.935  1045  0   0.937  671   0.642  79    158   83.67  #165
  4 GATAGAAACCAACCTGGC    TTACCTTAGGGATAACAG  53.6  2.0  47.7  27.0  9   7   GG   1511  0   0.943  1057  0   0.948  654   0.619  139   218   143.67  #165
```

Strict Primer Algorithm

E

ATACGGCTACTAACT
ATACGGCTACTAACT
ATACGGCTAGTAACT
ATT CGGCTACTAAAGT
ATT CGGCTACTAAAGT
ATT CGGCTACTAAAGT
ATT CGGCTACTAAAGT



Words of length L present
in at least S sequences of \mathbf{E}
 L : number (18-21)
 S : percentage (default=70)

$Lp(\mathbf{E})$

ATACGGCTACTAACT
ATT CGGCTACTAAAGT

Words of length L present in
at least S sequences of \mathbf{E} ,
and present in T sequences
of \mathbf{E} with no more than m
mismatches.
 T : percentage (default=90)
 m : number (1-3)

$Lp'(\mathbf{E})$

ATACGGCTACTAACT
ATACGGCTAGTAACT
ATT CGGCTACTAAAGT

Finds a space \mathbf{D} within the interval of amplified
sequence length $[l_{min}-l_{max}]$ and creates $Lp'(\mathbf{D})$. Pairs
 $Lp'(\mathbf{E})$ - $Lp'(\mathbf{D})$.

ATT CGGCTACTAAAGT - ATT CGGCTACTAAAGT

Bs
Bc

ecoPrimers

Lights:

- Computes B_c/B_s from amplified sequences.
- Constrains no mismatches in 3'-end of the primer.
- Pairs primers within an interval of barcode length.
- Considers ‘countersequences’.

Shadows:

- No degeneracy allowed. Mismatches are mismatches.
- Very taxonomy-constrained (EMBL, GB...) -> Whole genomes, no individual genes.

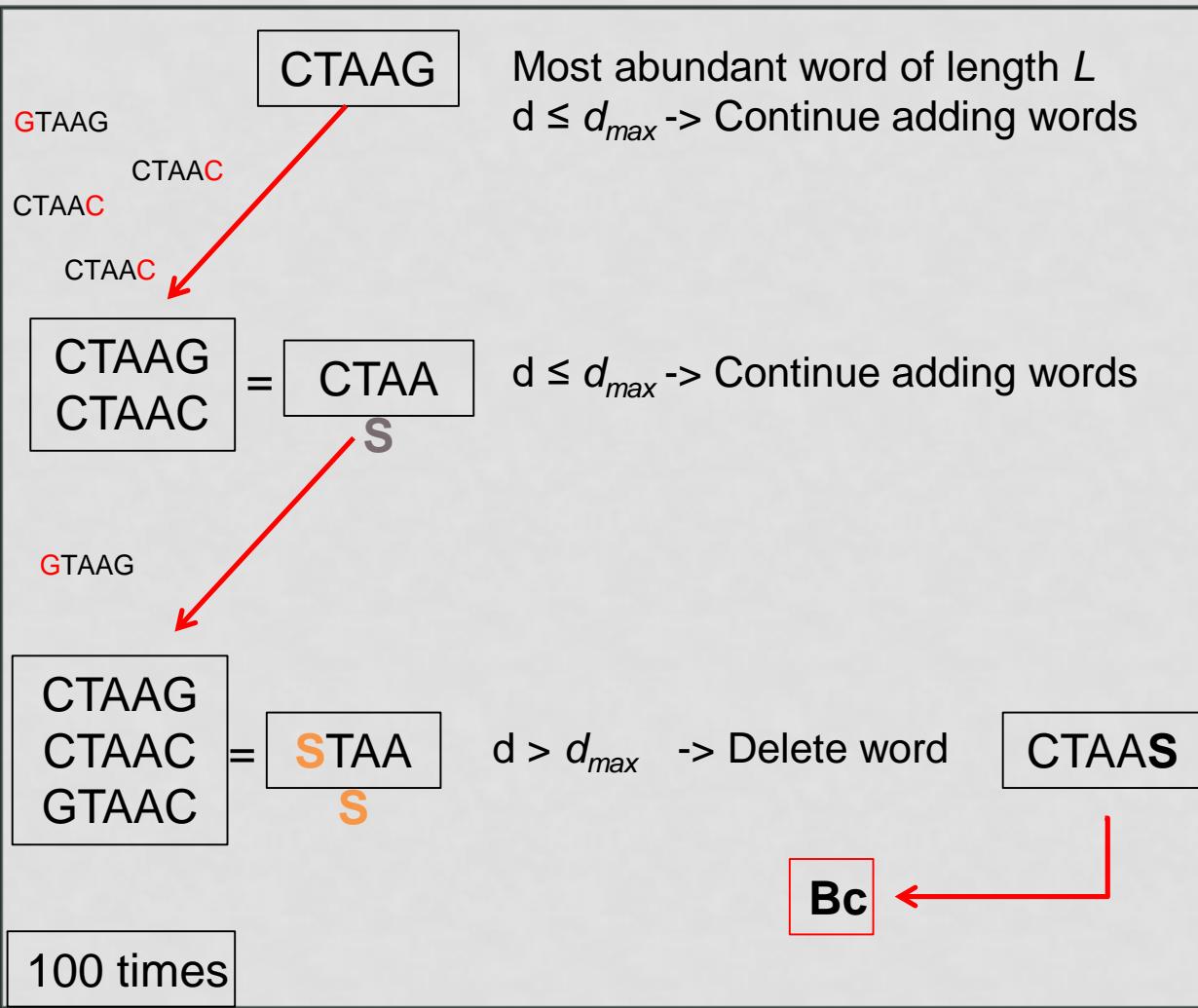
DegePrime (Hugerth *et al.* 2014)

- Developed at the SciLifeLab.
- Originally developed for 16S/metagenomics in prokaryotes.
- Perl based.
- Algorithm: Weighed Randomized Combination.
- Works over an alignment. It doesn't allow gaps, so the alignment should be modified in a program readable format (TrimAlignment.pl).
- Output has to be edited to be useful.

Weighed Randomized Combination

$d_{max} = 2$

ATTGGCTACTAAC
ATACGGCTACTAAC
ATACGGCTACTAAC
ATACGGCTAGTAAC
ATTGGCTACTAAAGT
ATTGGCTACTAAAGT
ATTGGCTACTAAAGT
ATTGGCTACTAAAGT



DegePrime

Lights:

- Allows degeneracy (not mismatches).
- Computes B_c .
- Gives measure of sequence diversity.

Shadows:

- No 3'-end constrain (but you can do it yourself).
- No pairing at length interval = No B_s index (but there are tools for doing it afterwards).
- Too much degeneracy when not needed (next slide).

ATTCGGCTACTAACT
ATACGGCTACTAACT
ATACGGCTAGTAACT
ATTCGGCTACTAAAGT
ATTCGGCTACTAAAGT

ATTCGGCTACTAA**G**T
ATTCGGCTACTAA**G**T

~~ATTENTION~~

ATACGGCTACTA**ACT**
ATACGGCTAGTA**ACT**

 AT{T/A}CGGC{T/C/G} TAA{G/C} T
d = 1x1x2x1x1x1x1x1x1x2x1x1x1x2x1 = 8

ATTCGGCTACTAAAGT
ATTCGGCTACTAAACT

ATACGGCTAGTAACT
ATACGGCTACTAACT

ATTCGGCTA{C/G}TAACT
 $d = 1 \times 2 \times 1 \times 1 \times 1 \times 1 \times x = 2$

LET'S DO SOME SCIENCE NOW

ecoPrimers

What do we need?

- Set of mitochondrial genomes downloaded from GenBank (gb format) ✓
- Taxonomy repository download from NCBI ✓
- Format the taxonomy into OBITools format ✓ “ncbi20150906”
- Format the genomes into OBITools database ✓ “sixlegs”

```
> cd ~/metabarcoding/  
> ecoPrimers -d sixlegs -e 3 -3 3 -l 50 -L 650 -r 6960 \  
-c > Insectsprimers.ecoprimers
```

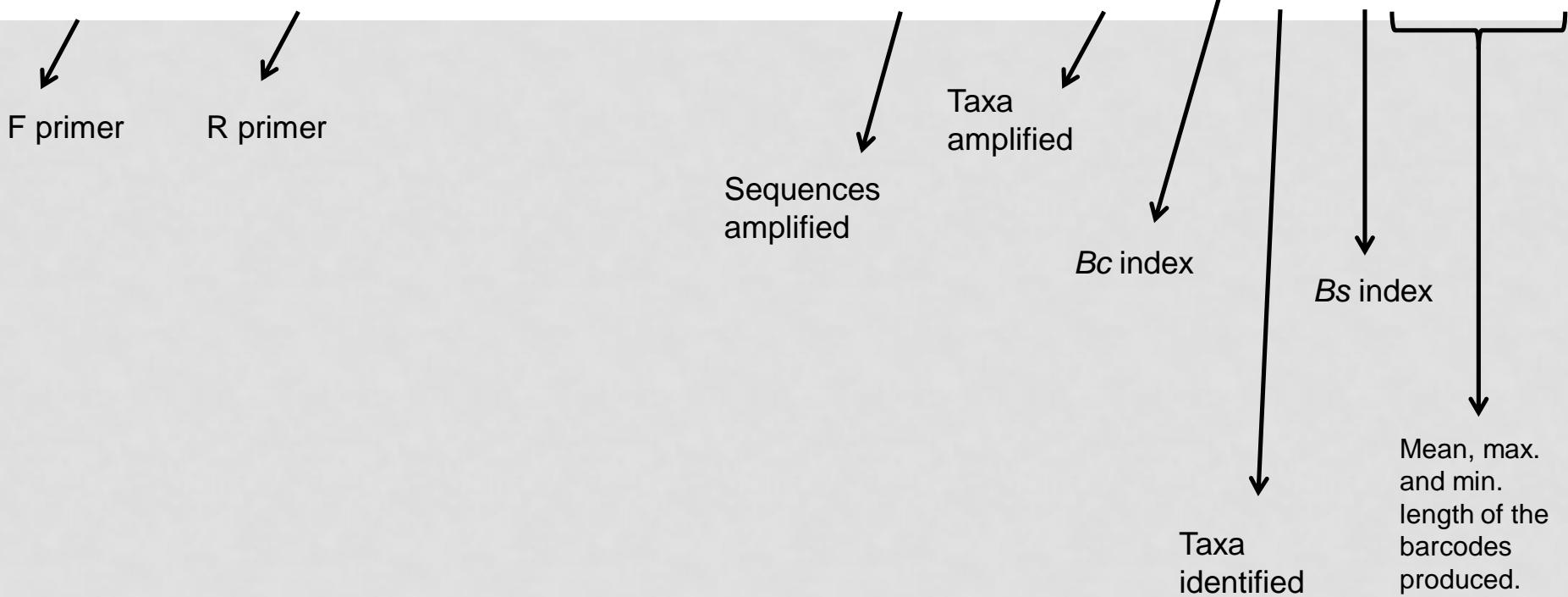
- d sixlegs: OBITools-format collection of genomes
- e 3: maximum number of mismatches in the second step
- 3 3: number of nucleotides of the 3' end constrained in a perfect match
- l 50 –L 650: minimum (*l*) and maximum (*L*) length of the potential barcode
- r 6960: NCBI taxid of the target group (when having other genomes too)
- c: sequences are circular (mtDNA)

> less Insectprimers.ecoprimers

```

# ecoPrimer version 0.3
# Rank level optimisation : species
# max error count by oligonucleotide : 3
#
# Restricted to taxon:
#   6960 : Hexapoda (superclass)
#
# strict primer quorum : 0.70
# example quorum : 0.90
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#
# database : sixlegs
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# amplifiat length between [50,500] bp
# DB sequences are considered as circular
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1  ATAGAAACCAACCTGGCT    TACCTTAGGGATAAACAGC  53.6  1.7   50.6  30.9   8   8   GG  1502  0   0.938  1048  0   0.940  824  0.786  137  216  141.67
2  GATAGAAACCAACCTGGC    TACCTTAGGGATAAACAGC  53.6  2.0   50.6  30.9   9   8   GG  1499  0   0.936  1046  0   0.938  822  0.786  138  217  142.67
3  ATAGAAACCAACCTGGCT    GACCTCGATGTTGGATT A 53.6  11.4  51.8  38.1   8   8   GG  1498  0   0.935  1045  0   0.937  671  0.642  79  158  83.67
4  GATAGAAACCAACCTGGC    TTACCTTAGGGATAAACAG  53.6  2.0   47.7  27.0   9   7   GG  1511  0   0.943  1057  0   0.948  654  0.619  139  218  143.67

```



DegePrime

What do we need?

- Set of mitochondrial genomes downloaded from GenBank (fasta format) ✓
- COI gen extracted from every genome ✓
- Alignment of the COI gen extracted ✓

```
> cd DegePrime

> perl TrimAlignment.pl -i COI_aligned.fasta -min 0.9 -o COI_trimmed

> perl DegePrime.pl -i COI_trimmed -d 12 -l 18 -o COIprimer

> less COIprimer
```

Pos	TotalSeq	UniqueMers	Entropy	PrimerDeg	PrimerMatching	PrimerSeq
20	1145	101	4.35512615524902	12	710	HAAYCATAARGATATTGG
21	1145	96	4.23929291093935	12	706	AAYCATAARGATATTGGH
22	1146	107	4.30799785685939	12	698	AYCATAARGATATTGGHA
23	1148	130	4.49977750333136	12	678	YCATAARGATATTGGHAC
24	1148	167	5.07112704195661	12	663	CATAARGATATTGGWACH
25	1151	224	5.55058302792518	12	589	ATAARGATATTGGWACHT
26	1151	223	5.54245740925846	12	590	TAARGATATTGGWACHTT
27	1151	216	5.35036312521185	12	627	AARGATATTGGWACHTTA
28	1151	215	5.34515272119099	12	628	ARGATATTGGWACHTTAT
29	1151	214	5.35138851544978	12	626	RGATATTGGWACHTTATA
30	1151	217	5.35568496151132	12	627	GATATTGGWACHTTATAY
31	1151	217	5.3372916008392	12	630	ATATTGGWACHTTATAYT
32	1151	213	5.31738112216666	12	633	TATGGWACHTTATAYTT
33	1151	223	5.52912247355897	12	599	ATGGGAACHTTATAYTTY
34	1151	284	5.90074965532906	12	552	TTGGAACHTTATAYTTYA
35	1151	283	5.90074965532905	12	552	TGGAACHTTATAYTTYAT
36	1151	320	6.1616857699409	12	505	GGAACHTTATAYTTYATT
37	1151	330	6.21183638827913	12	502	GAACHTTATAYTTYATTT
38	1151	329	6.2042021670173	12	503	AACHTTATAYTTYATTT

-i file: input file

-o file: output file

-min: proportion of gaps
“deleted”

-d: max degeneracy allowed

-l: length of the primer

```

> cat COIprimer > COIprimer.csv

> awk '{print $6, " ", $1, " ", $7, " ", $4, " ", $5}' COIprimer.csv > COIprimerfilter.csv

> echo "SeqMatched; Position; Sequence; Entropy; Degneracy" > COIprimer.csv

> sort -g -r -t" " -k1 COIprimerfilter.csv >> COIprimer.csv

> rm COIprimerfilter.csv COI_trimmed COIprimer

> less COIprimer.csv

```

SeqMatched	Position	Sequence	Entropy	Degneracy
959	686	ACAYTTATTYTGATTYTT	3.21773124654164	8
958	685	AACAYTTATTYTGATTYT	3.22689142492553	8
958	684	CAACAYTTATTYTGATTY	3.22689142492553	8
958	683	YCAACAYTTATTYTGATT	3.42525029766891	8
953	681	TAYCAACAYTTATTYTGA	3.47090117155666	8
950	682	AYCAACAYTTATTYTGAT	3.49294077700492	8
923	714	GAAGTHTAYATTYTAATT	3.35452347323419	12
911	911	WGCHACWATAATTATTGC	4.34891234548245	12
909	692	ATTYTGATTYTTGGDCA	3.92213973138692	12
909	691	TATTYTGATTYTTGGDC	3.92040362027581	12
908	910	CWGCHACWATAATTATTG	4.36594356035277	12
897	280	TAAATAAYATAAGHTTYT	3.50406186145596	12
896	281	AAATAAYATAAGHTTYTG	3.51302350097954	12
889	909	TCWGCHACWATAATTATT	4.5084895283526	12
879	689	YTTATTYTGATTYTTTGG	3.52505680452662	8
879	688	AYTTATTYTGATTYTTTG	3.52505680452662	8
879	687	CAYTTATTYTGATTYTTT	3.52505680452662	8
879	182	HATAATTTTYYTTYATAGT	3.74491098498773	12

ecoPCR

What do we need?

- Set of mitochondrial genomes downloaded from GenBank (gb format) ✓
- Taxonomy repository download from NCBI ✓
- Format the taxonomy into OBITools format ✓ “ncbi20150906”
- Format the genomes into OBITools database ✓ “sixlegs”
- Primer pair ✓ HATAATTTYTTYATAGT AARAATCARAATAARTGT
- OBITools package ✓

```
> cd ../  
  
> ecoPCR -d sixlegs -e 0 -l 50 -L 500 HATAATTTYTTYATAGT \  
AARAATCARAATAARTGT > COIpcr.ecopcr
```

- d sixlegs: OBITools-format collection of genomes
 - e 0: maximum number of mismatches primer-sequence
 - l 50 –L 650: minimum (*l*) and maximum (*L*) length of the amplified barcode
 - HATAATTTYTTYATAGT AARAATCARAATAARTGT: Forward and Reverse primers*
- *it doesn't matter the order of the primers

> less COIpcr.ecopcr

```
@ecopcr-v2
#
# ecoPCR version 0.2
# direct strand oligo1 : HATAATTTTYYATAGT ; oligo2c :
# reverse strand oligo2 : AARAATCARAATAARTGT ; oligo1c :
# max error count by oligonucleotide : 0
# optimal Tm for primers 1 : nan
# optimal Tm for primers 2 : nan
# database : sixlegs
# amplifiat length between [50,500] bp
# output in superkingdom mode
# DB sequences are considered as linear
#
FJ171325 | 16036 | 279481 | species | 279481 | Polystoechotes punctatus | 279480 | Polystoechotes | 279479 | Polystoechotidae
2759 | Eukaryota | D | TATAATTTTTTTATAGT | 0 | nan | AAAATCAAATAAAGT | 0 | nan | 486 | TATACCTATTGTTATTGGAGGATTTGGAATTGATTAGTTCC
TTAACATAGCAGCACCTGATAGCTTCCCACGAATAAAATAATAAGTTTGAAATTACCTCCCTTACTCTTTATTAGCATCAAGTATAGTTGAAAGAGGGCTGGTACAGGATGAACGTCTATCCACCTCTTCAGGAATTGGCTATGCAGGAGCTCTGTTGATTAGCAATTTCAG
TTACATTAGCCGGTATCATCGATTTAGGTGCTGTAATTAACTGTAATTAAATACGTTTACATACATAACTTTAGACCGAATACCTTATTGTTATGACTGTTTACAGCTTATTAACTTTATCTTACCTGTTGAGCTTACAATACTCTTACTGATCGTAA
TTAAATACATCATTTTGACCCCTGCTGGAGGAGGTGATCTTATATCA | Polystoechotes punctatus mitochondrial, complete genome
FJ171324 | 15877 | 559169 | species | 559169 | Ascaloptynx appendiculatus | 559167 | Ascaloptynx | 146494 | Ascalaphidae
2759 | Eukaryota | D | TATAATTTTTTTATAGT | 0 | nan | AAAATCAAATAAAGT | 0 | nan | 486 | AATACCTATTGTAATTGGTGGATTGGAAATTGATTAGTTCC
ACTTATACAGCCACCCAGACATAGCTTCCCACGAATAAAATAATAAGTTTGATTATTACCTCCCTTACATACACTTCTGCTGCTCATGCTGCAAGAGGGCTGGGACAGGTTGAACAGTTACCCCCCTTACGCTGGAATTGGCTATGCAGGTGCTCTGTTGACTTAGCCATTTCAG
TTACATTAGCTGGGTATCTCAATTAGAGGCTGTTAATTAACTGACTTTCTTATATAACACTTGATCGAATAACTTATTGTTGATCAGTTTACAGCAATTAACTTACTATATTACAGTTTACCTGAGGTGCAATTACTATATTAACTGATCGAAA
TCTAAATACATCATTTGACCCAGCAGGAGGTTGGAGACCCAATTAAATCA | Ascaloptynx appendiculatus mitochondrial, complete genome

F primer
R primer
Length of the barcode
Sequence of the barcode
```

```
> obitoools
```

```
> ecotaxstat -d sixlegs -r 6960 COIpcr.ecopcr
```

```
COIpcr.ecopcr 100.0 % |#####\ ] remain : 00:00:00
rank           ecopcr      db    percent
class          4           4    100.00
family         190        298   63.76
genus          461        763   60.42
infraclass     3           3    100.00
infraorder     19          21   90.48
kingdom        1           1    100.00
order          22          29   75.86
parvorder      2           2    100.00
phylum         1           1    100.00
species        673        1115  60.36
species group  14          18   //./8
species subgroup 8          13   61.54
```

```
> ecotaxspecificity -d sixlegs -e 14 COIpcr.ecopcr
```

-e 14: number of base errors to be considered the same species for determination = 0.03 of the barcode's length -485- (species identification threshold=97%)

```
Alignment : 0959 x 0982 -> 4604 99.5 % |#####\ ] remain : 00:00:00
rank           taxon_ok  taxon_total  percent
order          20         22    90.91
infraclass     3          3    100.00
superfamily    81         83    97.59
parvorder      2          2    100.00
species group  14         14    100.00
superkingdom   1          1    100.00
kingdom        1          1    100.00
phylum         1          1    100.00
infraorder     19         19    100.00
subfamily      210        213   98.59
class          4           4    100.00
species        593        673   88.11
superorder     1           2    50.00
suborder       20          21   95.24
```

```
> exit
```